



Protocol for DNA purification from formalin-fixed paraffin-embedded tissue

This protocol is for use of a QuickPick™ kit together with a QuicPick™ magnetic tool.

1. Cut a small section (10 - 20 mg) of the formalin-fixed paraffin-embedded tissue and manually remove any excess of paraffin as much as possible. Place the tissue in a 2 ml tube.
2. Add 1.2 ml Xylene to the sample and vortex vigorously.
3. Centrifuge at full speed for 5 min at room temperature.
4. Carefully remove the supernatant by pipetting. Do not remove the pellet.
5. Add 1.2 ml ethanol (96 - 100 %) to the pellet, and mix gently by vortexing.
6. Centrifuge at full speed for 5 min at room temperature.
7. Carefully remove the ethanol by pipetting. Do not remove the pellet. Repeat steps 5-7 once.
8. Incubate the tube containing the sample (with the cap open) at 37°C for 10 minutes until the ethanol has evaporated.
9. Immediately resuspend the tissue pellet in 50 - 200 µl TE Buffer. Homogenize and process the tissue according to the protocol described in the QuickPick™ SML gDNA kit insert.

Table 1. Reagent volumes for genomic DNA purifications

Reagent	Reagent volume per preparation			
	1.25 mg	2.5 mg	5 mg	10 mg
Sample amount				
Lysis Buffer	25 µl	50 µl	100 µl	200 µl
Proteinase K	2.5 µl	5 µl	10 µl	20 µl
Binding Buffer	62.5 µl	125 µl	250 µl	500 µl
Magnetic Particles	2 µl	4 µl	8 µl	16 µl
Wash Buffer 1	2 x 125 µl	2 x 250 µl	2 x 500 µl	2 x 750 µl
Wash Buffer 2	125 µl	250 µl	500 µl	750 µl
Elution Buffer	5 - 25 µl	10 - 50 µl	25 - 100 µl	50 - 200 µl